

Nashi

Sampling Notes

The nutritional status of nashi trees is monitored using soil tests and plant analysis. Annual monitoring is important to help sustain optimum levels and avoid nutritional disorders. If disorders do occur, rapid diagnosis is necessary to assist correction.

The standard leaf sampling period is mid to late summer. Data is also available for sampling in late spring if this should be necessary.

Leaf - Early Season

Sampling Time: Mid to late November.

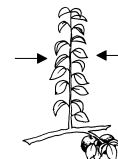
Plant Part Youngest mature leaf (blade & petiole).

Collect From: Mid portion of the current season's non-fruiting laterals (extension growth), taken at shoulder height.

Quantity per Sample: 4 representative leaves from the periphery of each of 15 - 25 trees.

Recommended Tests: Basic Plant (BP).

Comments:



Leaf - Mid Season

Sampling Time: January and February.

Plant Part Youngest mature leaf (blade & petiole).

Collect From: Mid portion of the current season's non-fruiting laterals (extension growth), taken at shoulder height.

Quantity per Sample: 4 representative leaves from the periphery of each of 15 - 25 trees.

Recommended Tests: Basic Plant (BP).

Comments: To help diagnose an obvious problem, leaves showing the first signs of the distinctive symptoms should be collected as soon as abnormalities appear. If sampling outside the normal sampling time it is useful to take a second sample of similar, healthy leaves from nearby unaffected trees for analysis as a comparative standard.



Soil

Sampling Time: Prior to crop establishment and annually at any time of the year, although autumn to early winter is recommended.

Core Depth 15cm.

Collect From: From the drip zone of the trees.

Quantity per Sample: 12 - 20 cores.

Recommended Tests: Basic Soil (BS).

Comments: Separate samples should be taken from blocks that differ in age, cultivar types, tree performance, soil types, topography and fertiliser history.

Where fertiliser has been broadcast, sample from the root zone of the trees. Where fertiliser has been banded, samples should only be taken from areas under the vines which have previously received fertiliser.

If the orchard has herbicide treated strips, then it is best if these are sampled separately from the grassed areas between rows. Quite different nutrient levels may exist between these two areas.

When sampling prior to orchard establishment, a 15 - 40 cm depth sample should also be taken, primarily to check the sub-soil pH.

Comments

Nitrogen, magnesium, calcium, manganese, zinc and boron nutrient disorders are the most common.

Adequate levels of nitrogen are required at blossoming and fruit set to ensure good fruit set.

Calcium related disorders, as well as magnesium deficiencies, may be induced by high levels of potassium that depress calcium and magnesium uptake. Foliar symptoms are not necessarily evident.

Manganese toxicity is more common than its deficiency, and is often related to low soil pH or poor drainage.

Boron is the most common deficiency, with symptoms of fruit shape distortion. Foliar symptoms are not necessarily evident.

It has been suggested that the soil potassium level should be 3-4% of the CEC. Calcium should occupy 70-80% of the CEC sites and magnesium 10-15%.

Pipfruit trees take up phosphorus efficiently, even in highly retentive soils. The suggested optimum Olsen P level is 30 or higher.

Pipfruit will grow within a soil pH range of 5.8 - 6.8. To minimise calcium disorders in fruit, a topsoil pH of 6.5 or greater is considered desirable.

References

Fertiliser recommendation for horticultural crops. HortResearch HortNET, 1997.

Blackmore, L.C; Searle, P.L and Daly, B.K. 1987. Methods for chemical analysis of soils. NZ Soil Bureau Scientific Report 80. NZ Soil Bureau, DSIR.

Nashi survey results, Nashi News - New Zealand Nashi Growers Association Inc. July, 1989.

Disclaimer

Normal Range levels shown as histograms in test reports relate specifically to the sampling procedure provided in this crop guide. The Normal Range levels in test reports and Comments provided in this Crop Guide are the most up to date available, but may be altered without notification. Such alterations are implemented immediately in the laboratory histogram reports. It is recommended that a consultant or crop specialist be involved with interpretations and recommendations.
